

Prevalence of insecticide resistance in field-collected populations of the German cockroach, *Blattella germanica* (Linnaeus) (Dictyoptera: Blattellidae) in Peninsular Malaysia

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Abstract: Thirty one field-collected strains of German cockroaches, *Blattella germanica* (L.), collected between February 1997 and March 2000 from various locations in Peninsular Malaysia were screened for insecticide resistance using topical application method against three commonly used insecticides (propoxur, chlorpyrifos and deltamethrin). A discriminating dose (LD₉₉) obtained from bioassaying a laboratory susceptible strain, was used to determine the prevalence of insecticide-resistant individuals among the field populations tested. Results indicated that both propoxur and deltamethrin resistance patterns were relatively similar, i.e. more than 50% of the strains screened show low to moderate level of tolerance, while chlorpyrifos resistance is still relatively low among the tested populations. Resistant gene frequencies estimated ranged between 0.01 to 0.78, 0.05 to 0.51 and 0 to 0.86 for propoxur, chlorpyrifos and deltamethrin, respectively.

INTRODUCTION

The German cockroach, *Blattella germanica* (L.) is an important urban insect pest in many parts of the world. Extensive usage of insecticides against this species has led to serious problems of insecticide resistance. Insecticide resistance in the German cockroach was detected as early as 1952 in Corpus Christi, Texas, USA (Heal et al., 1953) and subsequently numerous incidents of insecticide resistance in the German cockroach have been reported (Webb, 1961; Grayson, 1965; Cornwell, 1968; Bennett and Spink, 1968; Barson and McCheyne, 1979; Cochran, 1987; 1989; Horwood et al., 1991; Rust and Reiersen, 1991; Zhai and Robinson, 1991; Lee et al., 1996, 1997, 1999).

In Southeast Asia, the first document of broad spectrum insecticide resistance in the German cockroach was reported by Lee et al. (1996) in Malaysia, where twelve field-collected strains of German cockroach showed low to high resistance to carbamate insecticides (propoxur and bendiocarb), low resistance to organophosphate insecticide (chlorpyrifos) and low to moderate resistance to pyrethroid insecticides (cypermethrin, permethrin, phenothrin and deltamethrin), based on resistance ratios obtained from comparison of LD₅₀ values with that of a susceptible strain. Over the last few years, more incidents on insecticide resistance in the German cockroach in this region were detected and documented (Lee, 1997, 1998; Lee and Lee, 1998; Lee et al., 1997, 1999, 2000; Choo et al., 2000).

The ability to accurately detect the presence of resistant individuals in a population is crucial in a resistance management program. Dose-mortality regression method (topical application) was often reported to give higher resistance ratio values than time-mortality method (WHO glass jar contact) (Scott et al., 1986; Millio et al., 1987; Zhai and Robinson, 1992).

Discriminating dose (DD) is defined as the insecticidal dose that will kill all susceptible individuals. Insecticide bioassays utilizing a DD is more efficient in detecting the presence of resistant individuals in the population, than bioassays using a series of different concentrations or time intervals (Roush and Miller, 1986). Here, we report insecticide resistance in the field

populations of German cockroach by using discriminating dose. This method may not be able to elucidate the level of resistance in the German cockroach, but is extremely useful to detect potential development of insecticide resistance in the cockroach populations, even though the number of resistant individuals are only at an extremely small fraction of the population.

MATERIALS AND METHODS

A total of 31 field populations of the German cockroach were collected from various premises in Peninsular Malaysia from 1997 to 2000 to be screened for insecticide resistance. They were reared for

Table 1. Information on strains of field-collected populations of the German cockroach, *Blattella germanica* (L.) used in this study.

Strain	Collection site	City	Date of collection
BUSM	University cafeteria	Penang	20 Nov 1997
BBR	Hotel kitchen	Penang	11 Jan 1999
CIGO	Restaurant's kitchen	Kuala Lumpur	28 Dec 1998
COP	Hotel kitchen	Penang	21 Sept 1999
CP	Hotel kitchen	Penang	2 Nov 1998
CT	Hotel kitchen	Kuantan	18 Aug 1999
EMP	Cruise ship	—	2 Dec 1999
FBR	Hotel kitchen	Penang	20 Jan 2000
GCJB	Hotel kitchen	Johor Bahru	29 Dec 1998
GCPG	Hotel kitchen	Penang	29 June 1999
GL	Hotel kitchen	Kuantan	18 Aug 1999
GT	Hotel kitchen	Kuantan	3 Aug 1999
HUSM	University cafeteria	Penang	20 Nov 1997
IE	Express bus	—	24 May 1998
IHKL	Hotel kitchen	Kuala Lumpur	15 Oct 1999
Ita	Hotel kitchen	Penang	13 Dec 1999
KTM	Train	—	10 Mar 2000
LHFA	Restaurant's kitchen	Kuala Lumpur	25 June 1998
LHFB	Restaurant's kitchen	Kuala Lumpur	27 June 1998
Mal	Hotel kitchen	Kuala Lumpur	16 Dec 1998
Mar	Hotel kitchen	Penang	25 June 1998
ML	Hotel kitchen	Kuala Lumpur	13 Feb 1997
MT	Hotel kitchen	Penang	25 Jan 1999
PK	Hotel kitchen	Kuantan	17 Aug 1999
PRPG	Hotel kitchen	Penang	6 Jan 1999
Raja	Hotel kitchen	Kuantan	18 Aug 1998
Sedap A	Food court	Johor Bahru	5 Jan 1999
Selesa	Restaurant's kitchen	Johor Bahru	5 Jan 1999
Sun	Hotel kitchen	Penang	23 Nov 1998
Yao	Bakery	Penang	5 Dec 1998
ZT	Food court	Kuala Lumpur	28 Nov 1999

a generation under laboratory conditions of $26.5 \pm 0.5^\circ\text{C}$, $55 \pm 5.0\%$ relative humidity and a photoperiod of 12 : 12 (L : D) prior to insecticidal tests. For comparison, a susceptible strain, ICI, obtained from Zeneca Agrochemicals, U. K. was used as a standard in this study. Information on the field-collected strains are shown in Table 1.

Three commonly used insecticides, namely propoxur, chlorpyrifos and deltamethrin were tested against the field-collected insects. Technical grade propoxur (99.5%) (Bayer AG, Germany), chlorpyrifos (95.9%) (Dow Agrosciences Malaysia) and deltamethrin (96.1%) (Aventis Environmental Health, Malaysia) diluted in analytical grade acetone were used in this study. These insecticides were the three most common insecticides used in pest control operation in Malaysia. They are usually treated in the form of residual sprays.

LD₉₉ values for each insecticide were obtained by testing against a susceptible strain, ICI. A series of 5–7 concentrations of each insecticide, that caused 1–100% mortality, were used where 1.0 μl of the pre-determined concentrations of insecticides in acetone was applied topically onto the ventral mesothorax of CO₂-anesthetized male cockroaches. Each concentration was replicated three times with 10 insects per replicate. Control cockroaches were treated with acetone. Upon treatment, the insects were placed inside clean petri dishes and provided with food and water. Mortality was recorded at 48 hours post-treatment. Data were pooled and subjected to probit analysis (Finney, 1971) according to the procedures described by Robertson and Prestler (1992) using the POLO-PC program (LeOra Software, 1997).

Upon obtaining the LD₉₉ for each insecticide, all 31 strains of field collected populations were topically treated with 1 μl of LD₉₉ of each insecticide. Treated cockroaches were then placed into clean petri dishes and provided with food and water. Mortality of the cockroaches were recor-

ded at 48 hours post-treatment. Depending on availability, a total of 60–100 adult males were tested.

Resistant gene frequency was estimated by assuming the populations were in Hardy-Weinburg equilibrium (Falconer and Mackay, 1996) using the measured frequency of homozygous susceptibles in each strain. Dead cockroaches from treatment with discriminating dose were considered as homozygous susceptible individuals.

RESULTS AND DISCUSSION

The DD (LD₉₉) values obtained from the bioassays against the susceptible strain, ICI for propoxur, chlorpyrifos and deltamethrin were 3.027 $\mu\text{g}/\text{male}$, 2.540 $\mu\text{g}/\text{male}$ and 0.056 $\mu\text{g}/\text{male}$, respectively.

Overall results indicated that insecticide resistance in the field populations of German cockroach in Malaysia was generally prevalent (Fig. 1). Although propoxur has been reduced in usage in response to control failure, nevertheless, resistant genes have remained aloft in the population of the field strains, even under the absence of insecticide selection pressure. The discriminating dose of propoxur caused mortality range from 5.0 to 97.14% in the field populations tested (Table 2). This suggested low to high level of propoxur resistance. Propoxur-resistant gene frequency was estimated between 0.01 to 0.78. Earlier, Lee et al. (1996, 2000) reported that extensive usage and heavy reliance on propoxur have caused the development of high level of resistance to this insecticide in many populations of the German cockroach which were collected in the mid 1990s, in Malaysia. The results from this study suggested that propoxur resistance is still a very wide-spread problem, despite drastic reduction in its usage for several years.

Discriminating dose of chlorpyrifos killed between 24.0 and 90.0% of the field populations tested (Table 2, Fig. 1). Generally, resistance to chlorpyrifos was rela-

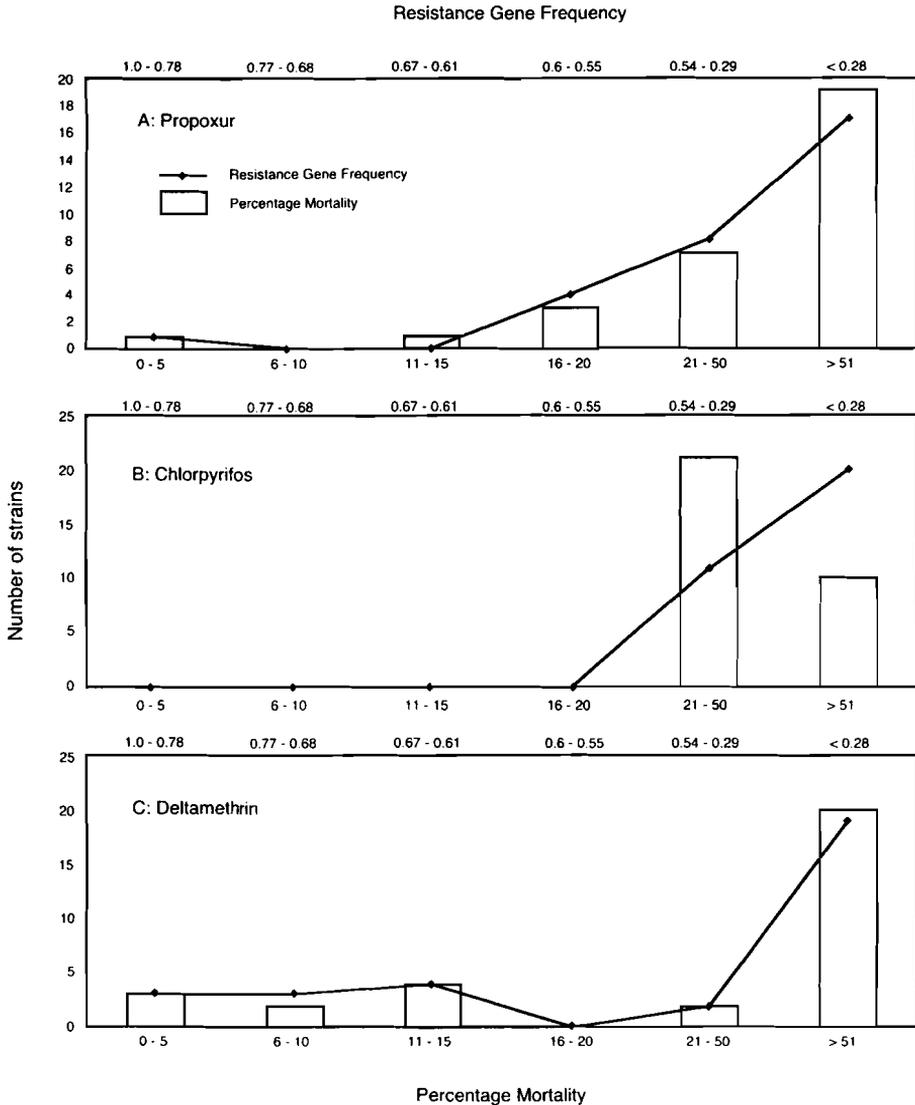


Fig. 1. Mortality of cockroaches treated with discriminating dosages of propoxur (A), chlorpyrifos (B) and deltamethrin (C). The number of strains are indicated by each bar adds up to 31. The line graph indicates frequency of resistant genes in the same mortality categories as the bar graph. The number of strains in the line graph also adds up to 31.

tively low with gene frequency estimated at 0.05 to 0.51. Only 29.0% of the total field populations tested carried at least 10% chlorpyrifos-resistant gene in the populations. Cochran (1995) reported that it was possible to detect insecticide resistance in a population of German cockroach if the frequency of resistant gene present in the population was at least 10%.

Cochran (1973) suggested that the discriminating dose technique could enable

the separation of the heterozygous and susceptible individuals, but not the resistant individuals. These findings corresponded well with the findings of Lee et al. (1996, 1999) where low resistance to organophosphate was recorded in field populations of the German cockroach. Cochran (1989) reported that high physiological resistance to organophosphate insecticides (chlorpyrifos and diazinon) was not detected although extensive usage of those in-

Table 2. Percentage mortality of the field-collected *Blattella germanica* after treatment with discriminating dose of propoxur, chlorpyrifos and deltamethrin.

Strain	Propoxur		Chlorpyrifos		Deltamethrin	
	<i>n</i>	% mortality	<i>n</i>	% mortality	<i>n</i>	% mortality
BUSM	100	96.00	90	73.33	100	94.00
BBR	60	80.00	100	43.00	100	60.00
CIGO	70	60.00	90	33.33	100	8.00
COP	100	55.00	90	44.44	100	13.00
CP	100	55.00	100	24.00	100	8.00
CT	100	20.00	100	67.00	100	5.00
EMP	100	5.00	100	31.00	100	30.00
FBR	100	30.00	100	58.00	100	45.00
GCJB	100	54.00	90	32.00	100	70.00
GCPG	100	40.00	90	71.11	100	75.00
GL	90	16.67	100	40.00	80	81.25
GT	100	15.00	100	55.00	100	63.00
HUSM	70	90.00	90	63.33	100	100.00
IE	100	54.00	100	90.00	100	65.00
IHKL	100	42.00	90	45.56	100	54.00
Ita	70	40.00	100	51.00	100	78.00
KTM	100	16.00	90	52.22	100	2.00
LHFA	100	78.00	100	54.00	100	94.00
LHFB	70	84.29	100	63.00	100	99.00
Mal	70	50.00	80	68.75	100	12.00
Mar	100	42.00	90	87.00	100	12.00
ML	100	68.00	100	67.00	100	15.00
MT	70	61.43	100	50.00	70	77.14
PK	100	48.00	100	48.00	100	53.00
PRPG	100	60.00	100	75.00	100	87.00
Raja	70	97.14	100	77.00	100	63.00
Sedap A	100	25.00	100	65.00	100	10.00
Selesa	100	90.00	100	76.00	90	88.89
Sun	100	70.00	100	65.00	100	76.00
Yao	100	86.00	100	78.00	100	5.00
ZT	100	17.00	100	55.00	100	60.00

secticides had been recorded, and therefore could still provide effective control of the German cockroach. Holbrook et al. (1999) also reported low to moderate resistance to chlorpyrifos in a more recent study. Our current findings showed that the use of chlorpyrifos to control the German cockroach in Malaysia could still provide good control as the frequency of resistant genes detected in field-collected populations was relatively low.

Results with deltamethrin showed a mortality range of 2.0 to 100.0% among the field-collected German cockroaches with an average mortality of 51.72% (Table 2, Fig. 1). Resistance to pyrethroids

have been numerous reported and incidents of control failure with this class of insecticide are on the rise (Gammon et al., 1981; Scott et al., 1986; Hemingway et al., 1993; Zhai and Robinson, 1992, 1996; Scharf et al., 1997; Lee, 1997; Lee et al., 1996, 1997, 2000; Lee and Lee 1998). This insecticide still display a good efficacy against the German cockroach where 64.5% of the field strains fall in the range mortality rates of >51%. Estimation of resistant gene of the field populations showed that the majority of the cockroach populations have yet to develop serious resistance to this insecticide where >60% of the field strains exhibited frequency of

resistant gene of less than 0.28. The frequency of resistant gene ranged from 0 to 0.86. Nonetheless, if the usage of the deltamethrin continues indiscriminately, the rate of frequency of the resistant gene would possibly increase in magnitude.

The results obtained suggest that the phenomenon of insecticide resistance is very common among broad-spectrum insecticides since all tested populations showed resistance to all classes of insecticide used in this study. A sensitive method is therefore crucial for early detection of resistant individuals. The method used in this study was able to serve this purpose relatively well. However, caution should be taken into account when dealing with the establishment of the discriminating dose because any error could affect the results of the detection of resistance to insecticides where susceptible strains might be wrongly treated as resistant strains. Another aspect that should be taken into consideration is the selection of susceptible strain to establish the discriminating dose. A clear distinction between the occurrence of natural tolerance in a population and the resistant genes has to be determined to avoid confusion when dealing with the level of insecticide resistance.

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